54th Annual Illinois Corn Breeders' School March 5-6, 2018



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Crop Sciences college of agricultural, consumer & environmental sciences



54th Annual Illinois Corn Breeders' School Program

March 5—6, 2018

At the I Hotel, 1900 South First Street, Champaign, Illinois

<u>Monday, March 5, 2018</u>

7:15-8:15	Registration and Continental Breakfast
Morning Sess Chair: Tony	sion: Breeding Processes Studer
8:15—8:30	Martin Bohn, University of Illinois Welcome and Introductory Remarks
8:30-9:15	Thomas Widiez, University of Lyon, France
9:15-10:00	Martha Willcox, CIMMYT25 <i>Title: Use of Landrace Maize in Plant Breeding at CIMMYT</i>
10:00-10:30	BREAK
10:30-11:15	Mark Mikel, University of Illinois 49 <i>Title: Progenitor lineages within proprietary dent corn germplasm</i>
11:15-12:00	Jenna L. Hoffman, Breeding Digital Phenomics and Statistics Lead, Monsanto 63 <i>Title: Reimagining our Fields</i>
12:00-1:15	Lunch
Afternoon Se Chair: Steve	ssion: Moose
1:15-2:00	William Gordon-Kamm, DuPont Pioneer69Title: Cereal transformation at DuPont Pioneer—meeting future demands for genome modification
2:00-2:45	Bing Yang, Iowa State70Title: Genome editing of maize
2:45-3:15	Break
3:15-4:00	Nicholas Heller, University of Illinois Graduate Student85Title: Response to selection in the ILTSE and a population of epigenetics NILS
4:00-6:00	Graduate Student Poster Session

54th Annual Illinois Corn Breeders' School Program



March 5-6, 2018 At the I Hotel, 1900 South First Street, Champaign, Illinois

Monday Evening: Chair: Steve Moose

5:30-6:30	Social Hour	
6:30-7:30	Dinner in the Chancellor Room	
7:30-8:15	Speaker: Matthew Hudson, Professor of Bioinformatics, Department of Crop Sciences, University of Illinois <i>Title: CS squared: Crop Science x Computer Science</i>	93

Tuesday, March 6, 2018

7:30-8:00 Registration and Continental Breakfast

Morning Session: Optimizing Corn Performance Chair: Martin Bohn

8:00-8:45	Dean Riechers, University of Illinois <i>Title: New Developments in Herbicide Resistance and Management Strategies for</i> <i>Waterhemp and Palmer Amaranth</i>	104
8:45-9:30	William L. Rooney, Texas A&M <i>Title: Validation and Implementation of Unmanned Aerial Systems in a</i> <i>Sorghum Breeding Program</i>	121
9:30-10:00	Break	
10:00-10:45	Maria Salas-Fernandez, Iowa State Title: A fieldp-based high-throughput phenotyping system for tall crops	122
10:45-11:30	David Hubert, BASF <i>Title: Phenotying for Fungal Resistance in Corn</i>	123
11:30-11:35	Closing Remarks	
List of Partici	pants	148

How to make maize seeds that look "not like dad": new insights in double fertilization and prospects for novel breeding tools.

Thomas Widiez, University of Lyon, France

Abstract:

Mixing male and female genetic information during sexual reproduction is considered as key to the evolutionary success of higher eukaryotes and is the basis of plant breeding. Sexual reproduction in flowering plants involves double fertilization, characterized by two separate fusion events between the male and female gametes. A maize line first reported in 1959 deviates from this classic pattern. Crosses using pollen from this so-called haploid inducer line, trigger the development of the egg cell into a haploid embryo with only the maternal genome, a process known as *in vivo* gynogenesis. Derivatives of this maize haploid inducer line have become the preferred tool of numerous maize breeding companies, because it can produce perfectly homozygous plants in only 2 generations instead of 5 to 8 in classical breeding schemes.

Fine mapping restricted a major QTL responsible for gynogenesis in maize to a zone containing a single gene coding for a patatin-like phospholipase A, which was named *NOT LIKE DAD (NLD)* because haploid embryos do not have paternal contribution. In all surveyed haploid inducer lines *NLD* carries a 4 pb insertion leading to a predicted truncated protein. This frameshift mutation is responsible for haploid induction as complementation with wildtype *NLD* abolishes the haploid induction capacity. Translational NLD::citrine fusion protein localizes to the sperm cell plasma membrane. In *Arabidopsis* roots, the truncated protein is no longer localized to the plasma membrane, contrary to the wildtype NLD protein. In conclusion, an intact sperm-specific phospholipase is required for successful sexual reproduction and its targeted disruption may allow establishing powerful haploid breeding tools in numerous crops.

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Laboratory "Reproduction & Développement des Plantes "



Who are we?

The "Seed development" team



Post-doc

PhD Student

Master Student

Gwyneth INGRAM Audrey CREFF Sophy CHAMOT

Douglas PYOTT

Jeanne LOU

Angelo GAITI

	endosperm
m	embryo
1	·

Peter ROGOWSKY Nathalie DEPEGE-FARGEIX Thomas WIDIEZ



Anne-Charlotte MARSOLLIER Laurine GILLES Nicolas DOLL

Post-doc PhD student PhD student



Maize transformation platform:

Ghislaine GENDROT Christelle RICHARD Edwige DELAHAYE

Main projects within "Seed dev"

Signaling/communication between seed compartments



Maize

Mechanism involved in haploid embryo induction



How to make Kids that Look "NOT LIKE DAD"?

✓ Context:



How to make Kids that Look "NOT LIKE DAD"?

✓ Context:



How to make Kids that Look "NOT LIKE DAD"?

✓ Context:



How to Make Maize Seeds that Look "NOT LIKE DAD"?

✓ Context:



Cross with inducer as male

How to Make Maize Seeds that Look "NOT LIKE DAD"?

✓ Context:



How to Make Maize Seeds that Look "NOT LIKE DAD"?

✓ Context:



in vivo gynogenesis: Production of maternal haploids via a male inducer

How to Make Maize Seeds that Look "NOT LIKE DAD"?

✓ Context:



✓ History of maize inducer lines :



Lashermes and Beckert 1988, Chalyk, 1994, Shatskaya et al. 1994, Eder and Chalyk 2002, Röber et al. 2005, Barret et al. 2008, Kebede et al. 2011...





The reasons to study in vivo haploid induction:

- Useful tool in maize breeding programs
- > Tool to understand plant reproduction (Double fertilization process)

The reasons to study in vivo haploid induction:

- Useful tool in maize breeding programs
- > Tool to understand plant reproduction (Double fertilization process)

* "Normal" double fertilization:



Gilles et al., Current Biology 2017

The reasons to study in vivo haploid induction:

- Useful tool in maize breeding programs
- Tool to understand plant reproduction (Double fertilization process)



✤ In vivo haploid induction (gynogenesis):





Pollen from inducer line



- 1- Identify the genetics behind maize haploid inducers
- 2- Understand molecular/cellular mechanisms
- **3-** Development of **breeding tools**

Fine based mapping of "ggi1" QTL

Barret *et al.*, 2008 \rightarrow *ggi1* (*gynogenesis inducer 1*) the main QTL for haploid induction



Fine based mapping of "ggi1" QTL

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Fine based mapping of "ggi1" QTL

Barret *et al.*, 2008 \rightarrow *ggi1* (*gynogenesis inducer 1*) the main QTL for haploid induction



→ PK6 gDNA region identified thanks to Pac-Bio sequencing of BACs



Fine based mapping of "ggi1" QTL





Identification of "NOT LIKE DAD" gene = NLD

NLD

Identification of "NOT LIKE DAD" gene = NLD



Article

Published online: February 22, 2017



Loss of pollen-specific phospholipase NOT LIKE DAD triggers gynogenesis in maize

Laurine M Gilles^{1,2}, Abdelsabour Khaled^{1,3}, Jean-Baptiste Laffaire², Sandrine Chaignon¹, Ghislaine Gendrot¹, Jérôme Laplaige¹, Hélène Bergès⁴, Genséric Beydon⁴, Vincent Bayle¹, Pierre Barret⁵, Jordi Comadran², Jean-Pierre Martinant², Peter M Rogowsky¹ & Thomas Widiez^{1,*} ¹



Identification of "NOT LIKE DAD" gene = NLD





- Two other parallel independent studies identified the same gene:

toi:10.1038/nat



MATRILINEAL, a sperm-specific phospholipase, triggers maize haploid induction

Timothy Kelliher¹, Dakota Starr¹, Lee Richbourg¹, Satya Chintamanani², Brent Delzer³, Michael L. Nuccio¹, Julie Green¹, Zhongying Chen¹, Jamie McCuiston¹, Wenling Wang¹, Tara Liebler¹, Paul Bullock² & Barry Martin¹?

CelPress

Molecular Plant Letter to the Editor

A 4-bp Insertion at *ZmPLA1* Encoding a Putative Phospholipase A Generates Haploid Induction in Maize

in Maize Chenxu Liu, Xiang Li, Dexuan Meng, Yu Zhong, Chen Chen, Xin Dong, Xiaowei Xu, Baojian Chen, Wei Li, Liang Li, Xiaolong Tian, Haiming Zhao, Weibin Song, Haishan Luo, Qinghua Zhang, Jinsheng Lai, Weiwei Jin, Jianbing Yan, Shaojiang Chen





Characterization of *NLD*

✓ *NLD* is expressed in mature anther



Is NLD expression pattern sporophytic (Anthers) or gametophytic (pollen)?

Gilles et al., EMBO J. 2017

Characterization of *NLD*

✓ *NLD* promoter is ON in mature pollen and OFF 3 days after pollination













Characterization of NLD





Confocal

Structured illumination microscopy

Gilles et al., EMBO J. 2017





Use of Landrace Maize in Plant Breeding at CIMMYT

Dr. Martha Willcox, Maize Landrace Improvement Coordinator International Maize and Wheat Improvement Center (CIMMYT)

Mexico, as the center of origin of maize, has contributed important genetic resources to germplasm banks, but native maize landraces, still sown as the majority of hectares of maize grown in Mexico also play a role as a reserve of genetic diversity still evolving through local farmer selection. CIMMYT has sought to use genetic resources, both ex situ and in situ, in improving lives of farmers and in providing information to breeders worldwide. As part of this effort the Seeds of Discovery Project, conducted both genotypic and phenotypic characterization on a large collection (over 4000) of maize landraces from the Breeders Core Collection of the Maize Germplasm Bank. This work has contributed information on allelic diversity but has also identified specific germplasm bank accessions that have used directly in breeding efforts for small farmers. Further work to improve landraces of marginalized communities in Oaxaca has focused on participatory breeding and agronomic improvements in areas where landrace maize is still the best option for farmers. This work has included farmer training, local diversity and selection studies, and cost benefit analysis of agronomic treatments. . During the course of this project, we have connected excess production of landrace maize by smallholder subsistence farmers of the project with culinary markets, opening a market that did not before exist. These market connections have increased prices of landrace maize to farmers and the rapid expansion of this market has created questions as to how to maintain the benefit of this market to smallholder subsistence farmers.





Use of Landrace Maize in Plant Breeding at CIMMMYT

Martha Willcox, CIMMYT Fernando Castillo Gonzalez, Colegio de Posgraduados Flavio Aragon Cuevas, INIFAP Humberto Castro Garcia, Universidad de Chapingo Leodegario Osorio Alcala, INIFAP

ICIMMYT.

- 60%+ of the maize hectares planted in Mexico are sown to Native Maize (traditional landraces).
 - Mexico is the center of origin of maize and conservation of native maize *in situ* is an international public benefit.
 - There are agroecological niches not served by hybrid programs.
 - Native Mexican Maizes are integral to the culture and culinary traditions of Mexico.
- Selling price of grain of Natives Maize is often higher than hybrid grain price.





Diversity of Culinary Uses





Seeds of Discovery





Yield and Agronomic Data Taken : All Locations

- Yield (field weight, grain and cob weight, moisture, number of ears)
- Plant Height and Ear Height
- Male and Female Flowering (50% of row)
- Stalk and Root Lodging



Tar Spot Complex

The symptom caused by Monographella maydis







MIn - Mean - Max TEPECI VANDEN-SALVAD -CLAVIL -COSTCR -DTRGLI -CANILL -CHANDE -HAITYE -EARCAR -CHAPAL CUBAAM CATETO CATETO PISANK AVMORO OLOTIL-CARIAC STCR01 TUSON PADENT PUYA FTCUBA CELAYA MCUBAN NORCAT RGDENT DTRGRU DENTAD

2.0

CONICO OLOTON TEHUA

DZITBA NALTEL

CIMMYT.

CRAVRG CAINGA RGWHDT TABLON

GUARIB

Selection Under Tar Spot Complex

- 2011B and 2012B Evaluation of TC under Seeds of Discovery
- 2013B Crosses between best Accessions and 4 CMLs.
- 2014A Backcross to Accessions Oaxa280 and Guat153
- 2014B Produce BC1S1s
- 2015B BC1S1 evaluated in Chiapas and Oaxaca (bulks 3 locations)
- 2016A Increase farmer voted BC1
- 2016B
 - BC1 Oaxa280/CML324//Oaxa280 in farmer fields in Santiago Yaitepec, Oaxaca

CIMMYT.



Tar Spot Disease Complex

Oaxa280//Oaxa280/CML 324

- Accession identified in field trials.
- Selected by farmers along with local farmer samples

CIMMYT.



S1 line from Germplasm Bank



CIMMYT.

Participatory Plant Breeding of Native Maizes in Marginalized areas of Oaxaca




Collaborators

M.C. Flavio Aragón INIFAP



Ing. Humberto Castro UACH

M.C. Leodegario Osorio INIFAP

CIMMYT.





Biodiversity, Participatory Plant Breeding and Agronomic Trials:

Community and Municipality	Altitude (masl)	Incline	Environment	Maize Races	Indigenous Groups	Percentage of the population in extreme poverty
El Sanjon & Rio Grande, Villa Tututepec	60	Flat	Arid Tropics	Conejo, Tuxpeño, & Olotillo	Mixtecos & Mestizos	20.94
Santiago Yaitepec, Juquila	1900	Hillslopes	Transition Zone	Comiteco	Chatinos	46.04
Santa Ana Zegache, Ocotlan	1600	Flat	Subtropics	Bolita	Zapotecos de Valle	29.16
Nduayaco & Jazmín Morelos, Santiago Apoala	2200- 2100	5 – 25%	Semi-arid Highlands	Chalqueño & Cónico	Mixtecos	49.3
Santa María Yavesia, Santa Maria Yavesia	2000 - 2100	5 - 35%	Humid Highlands	Cónico x Bolita, Elotes Occidentales, Serrano	Zapotecos	13.13



Participatory Improvement of Native Maize in Marginalized Areas of Oaxaca: San Antonio Nduayaco

> Altitude: 2300 masl Climate: Highland semi-arid Index of Extreme Poverty: 49% Migration Rate: 60% Soil Type: stony, red sandy loam, 2.3% OM, 20% slope, low in potasium, pH6.0 Biotic Stresses: White Grubs, Tar Spot, Turcicum Type of Maize: Chalqueño (cajete), Conico (temporal) Abiotic Stresses: Drought, Frosts

Planting Systems San Antonio Nduayaco



Density Plants/ha Plant Spacing Grain Yield (Kg/ha 14% Hum						
40.797	D1= 2 pl a 0.7 m	1494.875				
40,797	D2= 4 pl a 1.4 m	802.125				
-						
	Farmer Variety	Grain Yield (Kg/ha 14%	6 Hum			
	V4= Salvador	1503.00				
	V3= Juan	1461.25				
	V2= Isauro	840.25				
	V1= Andrés	789.50				
	Fertility	Grain Yield (Kg/ha 14%	6 Hum			
	F1= Balanced (soil test)	1862.375				
	F3= 50% of Balanced	1681.25				
	F2= Micorriza	672.125				
	F4= No Fertilizer	378.25				





CIMMYT.

Production Costs of Treatments

		No fertilizer	Fert. 50% (3.6 - 20-25 NPK)	Fert. 100% (7.2-	Mycrorrhiza
Land Preparation		rentinzer	20-23 NP KJ		Iviycionniza
Plowing \$350/team	4 teams/ha	1400	1400	1400	1400
Planting	· · · ·				
Landrace Seed \$7/kg	22 kg/ha	154	154	154	154
Labor \$150/day	12	1800	1800	1800	1800
Fertilization					
18-46-0		0	418.47	836.94	0
KCL			352.8	705.6	0
Micorriza		0	0	0	150
Application Labor	\$150/ person	0	150	300	0
Transport fertilizer	\$ 50/sack	0	85.5	171	0
Pest Control					
Seed Treatment		372.6	372.6	372.6	372.6
Cultivation \$300/team	2 teams/ha	600	600	600	600
Labor \$150/day	3	450	450	450	450
Manual weeding	4 laborers	600	600	600	600
Karate (2 aplic)	0.25 lt/ha		300	300	
Aplication Labor	\$150/person		300	300	
Harvest					
Labor	\$150/person	300	600	750	600
Shelling		300	600	750	600
Transport from field	A S		300	600	
Total (\$/ha)		5976.6	8483.37	10090.14	6726.6

Cost Benefit Analysis

			Cost Prod.	Value Prod.	Utility	
Farmer Seed Sample	Fertility Treatment	Yield (T/ha)	(\$/ha)	(\$/ha)	(\$/ha)	C/B Ratio
Sra. Antonia Alvarado	Per Soil Analysis (95-50-60 NPK)	5.09	11521.4	35,630	24109	3.09
Sra. Petra Pérez	Per Soil Analysis (95-50-60 NPK)	4.69	11521.4	32,830	21309	2.85
Sra. Antonia Alvarado	50% Recommended (47.5-25-30 NPK)	3.61	9399.4	25,270	15871	2.69
Sra. Petra Pérez	50% Recommended (47.5-25-30 NPK)	4.23	9399.4	29,610	20211	3.15
Sra. Antonia Alvarado	Mycorrhiza + 50% recommended (47.5-25-30 NPK)	3.82	9549.4	26,740	17191	2.80
Sra. Petra Pérez	Mycorrhiza + 50% recommended (47.5-25-30 NPK)	3.92	9549.4	27,440	17891	2.87
Sra. Antonia Alvarado	Farmer Fert. (50-0-0 NPK)	3.67	6761.8	25,690	18928	<u>3.80</u>
Sra. Petra Pérez	Farmer Fert. (50-0-0 NPK)	4.71	6761.8	32,970	26208	4.88
Sra. Antonia Alvarado	No Fertilizer	1.47	5928.0	10,290	4362	1.74
Sra. Petra Pérez	No Fertilizer	1.73	5928.0	12,110	6182	2.04
Local Price of Native Maiz	e \$ 7.0/kg					

Participatory Plant Breeding











Connecting Farmers to Culinary



Training Farmer Groups to Connect to Grain Cleaning Export Markets





Aflatoxin Testing





Commercialization Guidelines for Native Maize Landraces

Absent an authentication process for Native landraces, rapid expansion in the culinary market could cause inundation by nonlandrace maize and large commercial producers.

Experts in Native Mexican Maize were convened to:

- Define standards for distinguishing native maize
- Form a panel of experts to identify native maize
- Define types of farmers and communities that can be targeted.









Progenitor lineages within proprietary dent corn germplasm

Mark A. Mikel, Associate Director Biotech Center Institute for Genomic Biology, University of Illinois at Urbana-Champaign

During the past 40 years, the number of corn breeding programs has plummeted due to merger and acquisition, resulting in Pioneer Hi-Bred (PHI), Monsanto, and Syngenta owning ~95% of the U.S. PVP/utility patent registered lines. The first breeding cycles of proprietary germplasm began with publicly available germplasm and quickly transitioned to breeding within each company's own material. However, there are common progenitors across programs from using public/foundation germplasm and extensive breeding of PHI commercial hybrids during the early breeding cycles. For example, pedigree examination of PVP/patented lines registered since 2010 identify the two largest progenitors as B73 within the Stiff Stalk and PH207 within the Iodent heterotic family. Interestingly, B73 and PH207 contribute across proprietary germplasm pools. Among 461 non-Stiff Stalk inbreds registered since 2010, all (228 of 228) of PHI, 99% (167 of 169) of Monsanto, and 84% (54 of 64) of Syngenta lines are descendants of PH207. Among 391 Stiff Stalk inbreds registered since 2010, 95% (187 of 196) of PHI, 98% (143 of 146) of Monsanto, and 98% (48 of 49) of Syngenta lines are descendants of B73. As crosses between Stiff Stalk and Iodent heterotic families are the predominate commercial hybrid formulation, it is most likely that a hybrid grown in a farmers' field today in the heart of the U.S. corn belt is a cross between a B73 descendant and a PH207 descendant. Understanding the major lineages of commercial germplasm enables optimizing ex-PVP germplasm in forward breeding in predictable and sometimes novel ways.





Examples of Genetic Diversity in Vegetable, Fruit, and Field Crops



Transition from many to few companies

U.S. hybrid corn industry thirty years ago:

- · Era of "snow white" and the seven dwarfs and 100s of small companies
- ~400-500 U.S. located seed companies of all sizes retailing hybrid corn, soybean, and or small grains seed
 - ~40 had breeding program
 - ~100 multi-location hybrid evaluation in small plot or strip trials
 - nearly all had some form of hybrid evaluation (strip test or small plot)
- All but Pioneer Hi-Bred used foundation seed from Holden's, Illinois Foundation, or Brayton Seeds as sole source or supplementary germplasm

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U.S. corn % market share 1973 - 1983

Table 15—-U.S. market shares of corn seed by company ¹											
Company	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983
						Percent	t				
Pioneer	23.8	25.5	24.6	27.3	30.9	26.2	32.9	36.9	34.8	38.8	38.1
DeKalb ²	21.0	18.8	18.8	19.5	15.8	17.9	13.3	13.0	15.9	12.2	10.3
Asgrow	0	0	0	0	0	0	0	0	0	0	0
Funk ³	8.8	9.4	8.9	9.2	6.4	8.1	6.7	5.7	5.4	5.2	3.9
Trojan ⁴	5.9	5.1	6.8	5.6	4.2	5.4	3.8	2.0	0	0	0
Northrup-King ⁵	6.1	4.5	4.7	3.4	3.8	3.3	3.8	4.9	3.4	2.6	2.5
Zeneca/ICI	0	0	0	0	0	0	0	0	0	0	0
Cargill/PAG ⁶	4.8	6.8	3.9	3.5	4.1	4.6	3.3	4.7	5.6	5.4	4.2
Golden Harvest	0	0	1.8	2.4	2.5	3.1	2.9	1.3	3.2	2.3	2.6
Dow/Mycogen	0	0	0	0	0	0	0	0	0	0	0
Jacques/Agrigene	tics ⁷ 0	1.3	1.7	2	1.9	2.1	2.7	2.2	0	0	0
Other	29.6	28.6	29.8	27.1	30.4	29.3	30.6	29.3	31.7	33.6	38.4
Largest 8 firms	72.5	70.7	69.8	71.2	68.1	67.0	69.7	69.4	70.0	68.3	64.0
Largest 4 firms	59.7	58.8	59.1	61.6	57.3	55.6	56.7	60.5	59.5	59.1	54.9
Herfindahl index	0.1171	0.1159	0.112	0.1269	0.1049	0.1138	0.1354	0.1609	0.1501	0.1723	0.1604

from USDA Economic Research "The seed industry in U.S. agriculture" / AIB-786



Five Years of U.S. Seed Market Share

	C	ORN			
	2015	2014	2013	2012	2011
Monsanto	36.7	35.5	34.9	34.4	33.4
DuPont	34.6	34.7	36.0	36.1	35.8
Local & Regional Companies	10.5	11.1	11.0	11.8	13.0
AgReliant	7.1	7.0	6.7	6.5	6.6
Dow AgroSciences	6.1	6.0	5.3	5.2	4.7
Syngenta	5.0	5.7	6.1	6.0	6.5

https://www.agweb.com/article/seed-competition-heats-up-naa-sonja-gjerde/

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Bottom line of mergers and acquisitions is ...

_	Hybrid seed cost per acre					
_	1975	1995	2011	2017		
_	\$9	\$24	\$86	\$125		

What is the foundation of proprietary germplasm?

Input of public inbreds in first breeding
cycles of registered proprietary germplasm
(1980-1999)

Line	% Genetic contribution				
B73	12%				
B37	4%				
B14	4%				
A632	2%				
Mo17	6%				
Lancaster	6%				
Oh43	3%				
W117	1%				

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Footprint of PHI commercial hybrids in Monsanto and Syngenta inbreds registered 2010-present

		%GC in lines registered 2010-present			
PHI hybrid	# lines parent of	Monsanto	Syngenta		
PHI3901	15	1%	6%		
PHI3737	12	23%	10%		
PHI3394	7	1%	2%		
PHI3732	5	0%	0%		
PHI3751	4	0%	8%		
PHI3713	4	0%	0%		
PHI3378	4	2%	0%		
PHI3475	4	0%	0%		
PHI3540	3	0%	5%		
PHI3377	3	0%	0%		
PHI3358	3	0%	0%		
PHI3527	3	1%	0%		
PHI3535	2	1%	5%		
PHI3180	2	4%	0%		
PHI3293	2	0%	0%		
		34%	36%		

Also 2 lines developed from PHI hybrids: 3163, 3615, 3790, 3902, 3953, 3165, 3704, 3780,; and 1 line from 3147, 3160, 3369, 3720, 3978, 3162, 3195, 3199, 3245, 3603, 3769, 3558, 3861, 3921, and 3921, and LIZA.

Building block of first few 'PVP' breeding cycles

1980-1990: Most used germplasm (last breeding cross) of development of 271 registered Monsanto lines

	•	
Germplasm	Pedigree	Parent of
FBLL	5B2C-A / PB80	18
LH82	610 / LH7	17
B73	BSSS_C5	16
LH132	B73 *2 / H93	14
LH123	PHI3535	13
LH51	Mo17 isoline	13
3IIH6	PHI3737 (PHG29 / PHG47)	10
2FACC	4676A / PB80	8
LH74	A632 / B73	8
LH38	?A619 / L120?	7
Mo17	C103 / CI 187-2	7

1980-1990: Most used germplasm (last breeding cross) of development of 78 registered Syngenta lines

Germplasm	Pedigree	Parent of
B73	BSSS_C5	10
PHI3737	=PHG29 / PHG47	9
LH123	PHI3535	5
Mo17	C103 / CI 187-2	4
NP235	uk / A635	4
W117	643 / MINN13	4
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1980-1990: Most used germplasm (last breeding cross) of development of 252 registered PHI lines

Germplasm	Pedigree	Parent of		
PH207	PHB3BD2 / PHG3RZ1	17		
PHR03	PHT19 / PHG84	16		
PHJ40	PHB09 / B36	14		
PHW52	B73 / PHG39	13		
PHG39	PHA33GB4 / PHA34CB4	12		
PHP02	PHG44 / PHG29	12		
PHK29	PHB47 / PHAC54	10		
PHP38	PHG39 / PHK29	10		
PHG47	PH041 / MKSDTE	9		
PHR25	PHB83 / PH207	8		
PH595	50%FC, MYD, Oh07	7		
PHHB9	PHG86 / PHW52	7		

What is contemporary (2008-present) proprietary germplasm composted of?

			% Genetic Contribution				
Pedigee	Company/inbred	# hybrids	3IIH6	PH207	PHR03	Mo17	
	Monsanto Non-Stiff Stalk						
01INL1 / 17INI20	1285291	27	63%	33%			
1283669 / 1226218	CV197629	23	47%	18%			
I211988 / 2* I226218	CV651587	23	28%	11%			
01INL1 *2 / ASG5750	CV597869	21	56%	21%			
l119149 / l900429	CV805067	21	69%	30%		2%	
01INL1 / LH283	1226218	17	38%	14%			
	PHI Non-Stiff Stalk						
PHEDR / PH8JR	PH13JD	21		48%			
PHAVD / PH8CW	PHVAM	16		34%	19%		
PHVNV / PHNTV	PH1V5T	15		17%	31%	3%	
PH7DD / PH8JR	PHW2Z	15		51%			
PHACE / PHACV	PH128Z	10		37%	25%		
		10		11%	6%		

Most use	ed Stiff Stalk pare	nt in Mons	santo/PHI hybrids 2008 - present						
Pedigree	Company/Inbred	# hybrids	PHG39	90DJD28	2FACC	3AZA1	B73		
	Monsanto Stiff Stalks	-							
I900420 / I180421	CV995128	54			13%		28%		
PA2121 / 2* I294213	CV774864	32		38%	19%		35%		
01DHD10 / 90DJD28	1294213	20		50%	25%		47%		
1119135 / 1054029	CV483519	19			25%	38%	9%		
1325350 / 1119135	CV700979	19			13%	63%	5%		
1054029 / 1090372	CV789457	17		25%	25%	13%	19%		
	PHI Stiff Stalks								
PHE71 / PH7CH	PH12K5	20	27%				21%		
PH5WA / PH890	PHPAR	20	38%				31%		
PHAPT / PH890	PHV5W	18	38%				31%		
PH4GP / PH91V	PHW6G	17	38%				16%		
PHE0T / PH7CR	PH12SG	16	30%				15%		
PH4GP / PH714	PHF0D	15	31%				14%		
PH4GP / PH6WA	PHR1J	15	31%				14%		
PH09B / PH07D	PHCCW	13	31%				19%		

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Contribution among non-Stiff Stalk lines developed 2010-present

		Monsanto		PHI	Syngenta		
Line	% GC	Progeny	% GC	Progeny	% GC	Progeny	
Non-Stiff Stalk							
Mo17	1%	30% (52 of 169)	1%	14% (32 of 228)	6%	31% (20 of 64)	
PH207	18%	99% (167 of 169)	29%	100% (228 of 228)	16%	84% (54 of 64)	
PHR03			18%	72% (165 of 228)			
3IIH6 (carries 3/8 PH207)	38%	98% (165 of 169)					

Contribution among Stiff Stalk lines developed 2010-present

		Monsanto		PHI	Syngenta		
Line	% GC	Progeny	% GC	Progeny	% GC	Progeny	
Stiff Stalk							
B73	25%	98% (143 of 146)	17%	95% (187 of 196)	33%	98% (48 of 49)	
90DJD28	19%	67% (98 of 146)					
PHG39			26%	100% (196 of 196)			

In the U.S. Corn Belt the predominant hybrid 'formula' grown in a farmers' field is a Stiff Stalk female (B73 descendant) x lodent male (PH207 descendant)

ILLINOIS

Should we stick to the Stiff Stalk x non-Stiff Stalk paradigm?

-	Grain yi	in yield (bu/acre) half diallel 12 progenitors from 2 years (6 locations, ~12 reps)										reps)
		B73	PHG39	LH1	PHJ40	PH207	LH82	PHG47	Mo17	PHG35	PHG84	LH123
D	B73											
D	PHG39	166										
3	LH1	165	117									
3	PHJ40	159	146	130								
Ν	PH207	113	159	127	157							
0	LH82	179	146	139	152	147						
N	PHG47	150	156	149	158	139	162					
R	Mo17	175	146	156	139	153	153	135				
S	PHG35	183	161	149	138	170	150	169	137			
с С	PHG84	183	188	161	142	168	183	179	144	146		
3	LH123	180	170	152	167	152	178	164	141	171	181	
	PHZ51	175	172	173	160	152	167	165	151	158	126	166
					-BSS	NO	N-BSS	\$ →				

.

Hauck, Johnson, Mikel, Mahone, Morales, Rocheford, and Bohn. 2014. Crop Sci.

THE PLANTCELL

The lodent inbred PH207 genome

Draft Assembly of Elite Inbred Line PH207 Provides Insights into Genomic and Transcriptome Diversity in Maize^[OPEN]

Candice N. Hinsch,^{a, I}. Cony D. Hinsch,^p Alex B. Brohammer,^a Megan J. Bowman,^c Ilya Solfer,^a Omer Barad,^a Doron Snem Tov,^a Kobi Baruch,^a Fei Lu,¹ Alvaro G. Hemandez,⁹ Christopher J. Fields,⁹ Chris L. Wright,⁹ Klaus Koehler,⁷ Nathan M. Springer,¹ Edward Buckler,¹⁴ C. Robin Bueil,^{c,k} Natalia de Leon,^{1,m} Shawn M. Kaeppler,^{1,m} Kevin L. Childs,^{6,n} and Mark A. Mikel^{9,0}

Author enformation & Antick notice & Copyright and Upense Information &

ABSTRACT

Go to: 🐑

Intense artificial selection over the last 100 years has produced elite maize (*Zea mays*) inbred lines that combine to produce high-yielding hybrids. To further our understanding of how genome and transcriptome variation contribute to the production of high-yielding hybrids, we generated a draft genome assembly of the inbred line PH207 to complement and compare with the existing B73 reference sequence, B73 is a founder of the Stiff Stalk germplasm pool, while PH207 is a founder of lodent germplasm, both of which have contributed substantially to the production of temperate commercial maize and are combined to make eccess beterotic hybrids. Comparison of these two assemblies revealed over 2500 genes present in only one of the two constructions and 136 gene families that have undergone extensive expansion or contraction.





Identification of dispensable genes that are present in one genome but absent in the other through comparison of the two de novo assemblies and their specific annotation gene set

- 1,545 B73 specific genes that were present in B73 but absent in PH207 genome
- 2,042 PH207 specific genes present in PH207 but absent in B73 genome



Reimagining our Fields

Jenna Lynn Hoffman, Breeding Digital Phenomics and Statistics Lead, Monsanto

Valuable data layers can be derived from images sourced from satellites, manned aircraft, drones and smartphones. These data layers, when fused with other agronomic and product information, promise to deliver on insights specific to the acre. This will become the foundation for the combination of targeted breeding and personalized product placement. In this talk, I will highlight the new scientific insights that have resulted from this approach progress at Monsanto and Climate toward using this approach.





Reimagining our Fields

Monsanto Company Confidential

Illinois Corn Breeders' School Jenna Hoffman

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Shifting the Paradigm: Moving from measuring yield to understanding what is driving yield



Current Field Season

- Targeted data collection
- Trial assessment
- Contextual understanding





<u>Future Field Season</u> Full season data collection Embrace complexity Expand perspective



MONSANTO

Data Science as a Center of Excellence



Phenomics vision: Sense first, Sense global, Sense Remote, Sense smart

Genotype by phenotype specific sensing



MONSANTO



Cereal transformation at DuPont Pioneer – meeting future demands for genome modification.

William Gordon-Kamm Pioneer H-Bred International Abstract:

Keith Lowe, Emily Wu, Ning Wang, George Hoerster, Ajith Anand, Mauricio La Rota, Craig Hastings, Brian Lenderts, Mark Chamberlin, Maren Arling, Visu Annaluru, Candy Sweeney, Todd Jones & Bill Gordon-Kamm.

While transformation methods for monocot crops continue to improve, the process has remained constrained to a few genotypes per crop, and the methods have been slow and labor intensive, placing these methods beyond the reach of most academic labs. Recent progress in our labs is rapidly changing this situation for monocots. By focusing on the overexpression of the maize Babyboom (BBM) and Wuschel2 (WUS2) genes, we can routinely produce high transformation frequencies in numerous previously non-transformable maize inbreds. This was accomplished by altering the expression of our BBM and WUS cassettes in such a way that we can eliminate all callus steps and obtain transgenic T0 plants via direct germination of somatic embryos, making maize inbreds such as B73 and Mo17 easily transformable. Of even greater import to genome editing, this process is largely genotype independent and transgenic plants can be sent to the greenhouse in less than half the time of conventional methods. Another limitation for many monocots is the intensive labor and greenhouse space required to supply immature embryos for transformation. As a new alternative to immature embryos, we use BBM and WUS2 to recover transgenic events directly from either embryo slices from mature seed or leaf segments from seedlings in a variety of Pioneer inbreds, routinely recovering healthy, fertile T0 plants. Finally, we demonstrate that the maize BBM and WUS2 genes stimulate transformation in cereals.

T ILLINOIS Crop Sciences college of agricultural, consumer & environmental sciences Genome editing in maize

Bing Yang, Associate Professor Iowa State University, Ames, Iowa

Programmable nucleases (zinc finger nucleases, transcription activator-like nucleases, and CRISPR RNA guided Cas nucleases) have been successfully engineered to induce site-specific mutations at genomic loci in maize. The genome editing tools have significantly advance our basic understanding of gene function and engineering beneficial traits in maize. In my presentation, I will provide our experience in developing and utilizing TALENs (transcription activator-like nucleases) and CRISPR/Cas9 technologies for targeted mutagenesis in maize.



Genome editing in maize

Bing Yang <u>byang@iastate.edu</u> Iowa State University

Ames, Iowa 50011

Outline:

- Introduction of genome editing in plants
- An example of maize gene mutagenized by using TALEN technology
- Development and application of CRISPR/Cas9 in maize
- Conclusion


Plant genome editing requires transgenics







Major plant species targeted for gene editing with engineered nucleases

Plant	Mega- nuclease	ZFNs	TALENs	Cas9/gRNA
Arabidopsis	V	V	V	V
Canola		V		
Cotton	V			
Potato			V	V
Soy bean		V	V	V
Tobacco		V	V	V
Tomato			V	V
Barley			V	V
Maize	V	V	\checkmark	V
Rice			V	V
Sorghum				V
Wheat			V	٧

The Role of Genome Editing in Plant Biology and Agriculture

- Basic biology e.g., Functional genomics
- Gene/trait discovery e.g., SNP variations in gene expression and function
- Applied biology e.g., create novel germplasm with precise edits

Types of DNA Modifications with Genome Editing in Plant

- SNPs (single nucleotide polymorphisms
- Indels (Insertions/deletions)
- Large chromosomal deletions
- Insertion of gene or regulatory DNA
- Gene replacement

Benefits of Genome Editing in Agriculture

- Increase crop productivity and food production
- Increase resistance to plant pathogen and pest
- Increase tolerance to abiotic stress
- Better manage weeds
- Make healthier and more nutritious food



Structure of TAL effectors



- Repeat region determines the gene specificity
- Both NLS and AD are required for TALE activity

Yang et al. 2004 MPMI 17:1192-1200 Yang et al. 2000 PNAS 97:9807-9812 Zhu et al. 1999 Plant Cell 11:1665-1674





Plant Biotechnology Journal

Plant Biotechnology Journal (2015) 13, pp. 1002-1010



doi: 10.1111/pbi.12344

Heritable site-specific mutagenesis using TALENs in maize

Si Nian Char^{1,†}, Erica Unger-Wallace^{1,†}, Bronwyn Frame², Sarah A. Briggs¹, Marcy Main², Martin H. Spalding¹, Erik Vollbrecht¹, Kan Wang² and Bing Yang^{1,*}

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Received 3 November 2014; revised 18 December 2014; accepted 22 December 2014. *Correspondence (Tel +1 515 294 2968; fax +1 515 294 5256; email byang@iastate.edu) ¹These two authors contribute equally to this work.

Keywords: TAL effector nuclease, gene editing, targeted mutagenesis, maize, *Glossy2*.

Summary

Transcription activator-like effector nuclease (TALEN) technology has been utilized widely for targeted gene mutagenesis, especially for gene inactivation, in many organisms, including agriculturally important plants such as rice, wheat, tomato and barley. This report describes application of this technology to generate heritable genome modifications in maize. TALENs were employed to generate stable, heritable mutations at the maize glossy2 (gl2) locus. Transgenic lines containing mono- or di-allelic mutations were obtained from the maize genotype Hi-II at a frequency of about 10% (nine mutated events in 91 transgenic events). In addition, three of the novel alleles were tested for function in progeny seedlings, where they were able to confer the glossy phenotype. In a majority of the events, the integrated TALEN T-DNA segregated independently from the new loss of function alleles, producing mutated null-segregant progeny in T1 generation. Our results demonstrate that TALENs are an effective tool for genome mutagenesis in maize, empowering the discovery of gene function and the development of trait improvement.

TALEN construct and the target maize *Glossy2* sequences





Genetic segregation produces null segregants of edited *gl2* in the T1 generation





Glossy phenotype of TALEN-mutagenized glossy2 maize

TAL effector nucleases targeting the Glossy2 gene induce site-specific mutations that confer classic glossy phenotype. Water drops adhered to the surface of the mutant leaf (left) due to reduced epicuticular wax caused by loss of function of the *gl2* gene but not the wild type (right).

Plant Biotechnology Journal

Plant Biotechnology Journal (2016), pp. 1-12



doi: 10.1111/pbi.12611

An Agrobacterium-delivered CRISPR/Cas9 system for high-frequency targeted mutagenesis in maize

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Summary

CRISPR/Cas9 is a powerful genome editing tool in many organisms, including a number of monocots and dicots. Although the design and application of CRISPR/Cas9 is simpler compared to other nuclease-based genome editing tools, optimization requires the consideration of the DNA delivery and tissue regeneration methods for a particular species to achieve accuracy and efficiency. Here, we describe a public sector system, ISU Maize CRISPR, utilizing *Agrobacterium*-delivered CRISPR/Cas9 for high-frequency targeted mutagenesis in maize. This system consists of an *Escherichia coli* cloning vector and an *Agrobacterium* binary vector. It can be used to clone up to four guide RNAs for single or multiplex gene targeting. We evaluated this system for its



CRISPR/Cas9 system for targeted mutagenesis in maize



Cas9/gRNA continuously induces mutations in progeny plants



Table 1 Summary of CRISPR mutagenesis frequencies on four genes in maize Hi-II genotype

gRNA	Target gene	# bar+ callus line analysed	# Mutation+ callus line	% Mutation frequency	# Monoallelic mutant	# Diallelic mutant	# Mutation+ line regenerated
gAGO18a	ZmAgo18a	23	17	74	12	5	17
gAGO18b	ZmAgo18b	23	16	70	9	7	16
gAGO18a/b	ZmAgo18a	26	3	12	1	2	22
	ZmAgo18b		4	15	3	1	
	ZmAgo18a&18b		15	58	11 (18a), 10 (18b)	4 (18a),5 (18b)	
gA1/A4	a1	47	7	15	1	6	35
	a4		23	49	1	20	
	a1 & a4		7	15	0 (a1), 0 (a4)	7 (a1), 7 (a4)	

CRISPR/Cas can be used for multiplex targeting, e.g., producing up to 8 guide RNAs



Conclusion

- Engineered nucleases (ZFNs, TALENs, and guide RNA-directed Cas9) are promising genetic tools for genome editing in plants;
- Engineered TALENs are feasible for targeted mutagenesis in maize
- CRISPR/Cas9 is highly efficient to induce site-specific gene mutations in maize

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IOWA STATE UNIVERSITY

Response to Selection in the ILTSE and a population of epigenetic NILs

Nicholas Heller, Graduate Research Fellow, Department of Crop Sciences and Steve Moose, Professor, Department of Crop Sciences, University of Illinois at Urbana-Champaign

Advances in sequence technology have allowed incredible discoveries about the genomes of many organisms and elucidated relationships between gene and phenotype. These advances have allowed a deeper look into how variation in phenotype is created, selection for this variation is realized, and how phenotypes are inherited. However, many studies found that the heritability of many phenotypes is not fully explained by genomic DNA sequence, especially for quantitative, complex traits.

Here, I present some insights into the contributions to phenotypic response to selection in plants using a genetic system (utilizing the Illinois Long Term Selection Experiment, ILTSE), transgenic system (utilizing the red fluorescent protein driven by the *Floury2* promoter in the maize kernel), and an epigenetic system (utilizing variation created by the *mop1* mutation). Briefly, the ILTSE materials provide a unique resource because they have undergone continuous selection for the same trait for over 100 years and the last 50 years of seed is preserved. Reverse selection experiments are still underway to determine the plasticity of the populations' genomes after 50, 90, and even 100 years of forward selection. Finally, we utilize an inbred system to look more closely at the possibility that some of the response to selection is due to heritable, non-genetic factors.



Response to Selection in the Illinois Long-Term Selection Experiment and a Population of epigenetic NILs

Presented by Nicholas Heller

I ILLINOIS Crop Sciences GE of Agricultural.com



Department of Crop Sciences College of ACES, U of I Illinois Corn Marketing Board Corn Marketing Board Fellowship



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The Moose Laboratory

Crop Sciences College of Agricultural, Consumer, a environmental sciences **Past and Present**

Illinois Long-Term Selection Experiment



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The mop1 mutation





Barber et al., 2012

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Selection on a Reporter Phenotype – the Red Fluorescent Protein



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The Zein-RFP system is a worthy reporter for three reasons:

- 1) the phenotype (red seed) can be quantitatively measured through imaging techniques;
- 2) the alpha-zeins are responsive to nitrogen supply; and
- 3) alpha-zein genes are known to be sensitive to epigenetic regulation (Miclaus, *et al.*, 2011)

mop1-induced Phenotypic Variation

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Normal

Different 'Light Shows'





- Creation of epiNIL population
- Genetic similarity of epiNILs (to each other and to B73 control)
- Selected on RFP phenotype to create 34 'versions' of B73 (15 High RFP, 15 Low RFP, 4 Medium RFP) plus 4 lineages of the control B73:RFP

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Is the created variation heritable to hybrids?



Cross 14 inbred parents
to the epiNILs:

B73	Mo17	PH207
A632	PHZ51	LH82
PHG39	PHG84	PHJ31
LH1	PHJ33	IHP1
	NC350	ILP1

 Subsets of the ~500 hybrids grown over two years

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- Included select other control crosses between non-epiNIL inbreds
- Measured Height, Lodging, Grain Yield, Grain composition (Protein, Starch, Oil, etc.), and the RFP phenotype



CS squared: Crop Science x Computer Science

Matthew Hudson, Professor of Bioinformatics, Department of Crop Sciences University of Illinois at Urbana-Champaign

<u>Abstract</u>

The influx of data into Crop Sciences research and development, especially corn breeding, is becoming a flood. At the same time, the increase in the speed of computers (Moore's Law) is slowing down. In order to use the huge amounts of data that are being generated in Agriculture, new types of students and degree programs are needed. The University of Illinois is now offering a joint degree program in Crop Sciences and Computer Science, with the first students starting in Fall 2018. The need for this program and the curriculum will be described in the presentation.



& ENVIRONMENTAL SCIENCES

(CS)² Crop Science x Computer Science Matt Hudson University of Illinois

Science vs Malthus (1766-1834)

- Population, when unchecked, increases in a geometrical ratio, Subsistence, increases only in an arithmetical ratio.
- The power of population is so superior to the power in the earth to produce subsistence for man, that premature death must in some shape or other visit the human race.
- But
- The main peculiarity which distinguishes man from other animals, is the means of his support, is the power which he possesses of very greatly increasing these means.













How many of those people have wireless internet?



Computers also have a long history



Antikythera mechansim, ~100 BCE





Salamis Tablet, 300 BCE







Moore's law isn't going to fix the problem

- We need CS experts qualified to write better algorithms and apps specifically for agriculture
- We need to connect all those wireless data subscribers to their food supply
- We need to get farms connected in the way factories are (Buildings now have operating systems. Machines have their own networks)

The CS² undergrad program at U Illinois

- Joint degree offered by departments of CS and CPSC
- Students pay tuition at CS rate
- CS Core
- CPSC Core
- Equal weight

CS Core + Technical Track

- 7 Required CS classes at 100, 200, 300 and 400 level
- Includes data structures, algorithms, programming languages and compilers
- Technical track: Also systems programming, computer architecture, two additional 400 level CS classes from approved list

CPSC Core and other requirements

- Genetics, Biotech & Genetic Engineering, Data Science, Stats, Weed Science, Entomology, Plant Path, Crop Management, Plant Breeding, Genomics
- Plus 3x MATH courses, Probability and Stats for CS, CPSC Professional Development, Writing and Public Speaking, ACES 101 and usual Campus reqs (eg Foreign Language).

Enrollment

- First BS students start in Fall 2018
- We're expecting 20-25 per year
- Professional MS degree watch this space

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Thank you

- Fred Kolb, German Bollero (CPSC)
- Rob Rutenbar, Vikram Adve (CS)



and DNA s earch Ce ed and on w ays to treat and prevent plant, and and h

New Developments in Herbicide Resistance and Management Strategies for Waterhemp and Palmer Amaranth

Dean E. Riechers, Professor Department of Crop Sciences, University of Illinois at Urbana-Champaign

Abstract

Waterhemp (*Amaranthus tuberculatus*) and Palmer amaranth (*A. palmeri*) are problematic weeds in areas of the U.S. that produce corn, soybean, and cotton, mainly due to their competitive ability, outcrossing nature, genetic diversity, and resistance to multiple herbicide sites-of-action (SoA). Additionally, extended emergence and the ability to produce thousands of seeds per plant allow waterhemp and Palmer amaranth to quickly infest fields if proper preventative or control measures are not taken. Early season competition in corn has the largest effect on yield; the potential for up to 50% yield losses exists if weed control measures are not implemented before V6. Utilizing preemergence (PRE) herbicides to provide residual weed control in corn and soybean during early crop establishment is advantageous in limiting crop-weed competition and reducing the number of plants for postemergence (POST) control.

Waterhemp and Palmer amaranth populations on most farms have resistance to at least one SoA. However, previous research by our weed science group reported multiple resistances to HPPD inhibitors and other POST herbicides in a waterhemp population from central Illinois (MCR), as well as in a waterhemp population from Champaign County (CHR) exhibiting multiple resistances to HPPD inhibitors, atrazine, and auxin herbicides such as 2,4-D. CHR and MCR have also demonstrated variable control with different acetamide herbicides applied PRE. Collectively, these findings indicate that waterhemp and Palmer amaranth populations in the U.S. possess multiple mechanisms conferring complex cross- or multiple resistance patterns. Laboratory research at UIUC successfully determined underlying mechanisms and identified gene candidates conferring multiple resistances to mesotrione, topramezone, atrazine, imazethapyr and primisulfuron-methyl in MCR. Based on rapid atrazine metabolism, our recent research led to a unique diagnostic tool based on expression of a single *GST* gene to determine whether atrazine-resistant waterhemp possesses metabolic or SoA-based mechanisms. This *GST* can be used as a molecular marker to screen resistant waterhemp populations and, as technology advances, knocking out this *GST* could potentially reverse atrazine resistance. Research conducted at UIUC has revealed when and how resistance occurs in waterhemp and Palmer amaranth in an effort to gain insight into weaknesses that could be exploited for unique and innovative control measures. This new information is necessary to combat existing resistant weeds, prevent new resistant weeds from developing, and ultimately optimize crop yield.



Crop Sciences college of agricultural, consumer & environmental sciences



Outline of Presentation

- Biology of the dioecious amaranths = waterhemp and Palmer amaranth
- Genetic diversity favors development of herbicide resistant traits
- Integrated weed management systems based on research at UIUC




Waterhemp Management Guide – 1997



WATERHEMP MANAGEMENT IN AGRONOMIC CROPS

Aaron G. Hager Word Science Exter on Specialist. Department of Crop Sciences.

Loyd M. Wax Word Scientist, United States Department of Agriculture/Agricultural Research Service and Department of Crop Sciences, University of Illinois

F. William Simmons Associate Professor, Departu Sciences, University of Illing ent of Natural Resources and Envir

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ACES College of Agricultural, Consumer and Environmental Bolences

WATERHEMP MANAGEMENT IN ILLINOIS AGRONOMIC CROPS

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ur inte Amaranth, some basic identification of are provided in Table 1. For further in pigwood identification, consult an exc reed identification, consult an excellent guide loped jointly by Karage Sco Internet and A Construction of the Defendence of the Construction of t

ter, 16 Umberger Hall, Kansas State University, Man-hattan, KS 66506-3406.

WATERHEMP BIOLOGY

and Pal

WATERPENP BIOLOCY Market Market Market Menny are pigeword species faive to films, Historically, their distribution has faive to films, Historically, their distribution has the second second second second second function of the second second second second biasouri and lows. An increased presence of second second second second in central and castern Historical second seco

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Waterhemp (A. tuberculatus) Biology



C4 dicot and dioecious; often confused with Palmer amaranth (*A. palmeri*)

Can be differentiated by their female flowers and petiole:leaf length ratio

Waterhemp has evolved resistance to **six different** herbicide sites of action, including numerous *multiple-resistant* populations (and individual plants!)



- sound familiar?



... then turn left. It's the place with the weedy-looking corn."



Pratt et al. 1999.





Palmer amaranth Biology

- C4 dicot
- Dioecious male or female plants
- Inflorescence can be up to 1 meter long
- Produces 200 to 600,000 small seeds
- <u>Multiple emergence events</u> within season
- Distinguishing characteristics
 - long petioles and/or watermarks
- Reduction in corn yield 11-91% with a density of 0.5 to 8 plants per meter
 - 7-35% reduction when emerging after corn







Current Status

Resistance to herbicides from 6 Sites of Action

- Microtubule Inhibitors
 - Preemergence herbicides
- PS II inhibitors (atrazine)
- HPPD inhibitors
 - Only Amaranthus species have documented resistance so far...
- ALS inhibitors
- EPSPs (glyphosate)
- PPO inhibitors



Palmer amaranth confirmed in counties

Orange – counties with glyphosate-

resistant Palmer amaranth

Amaranthus palmeri

Distribution in Illinois

Multiple herbicide-resistant Amaranthus tuberculatus in east-central Illinois.

MERINAL Sites of action:

ALS inhibitors **HPPD** inhibitors Growth regulators (auxins) **PPO** inhibitors **PSII** inhibitors

Metabolic herbicide resistance in dioecious *Amaranthus*

HPPD inhibitors S-triazines (atrazine, simazine) ALS inhibitors



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(excised leaf assay for metabolism)



http://www.jove.com/video/53236/measuring-ratesherbicide-metabolism-dicot-weeds-with-an-excised-leaf





Illinois HPPD-inhibitor Resistant Population

<u>Year</u>	<u>Crop</u>	<u>Herbicides Applied POST*</u>
2003	Seed Corn	mesotrione + atrazine
2004	Seed Corn	mesotrione + atrazine
2005	Seed Corn	mesotrione + atrazine
2006	Seed Corn	topramezone + atrazine
2007	Seed Corn	topramezone + atrazine
2008	Seed Corn	tembotrione followed by mesotrione
2009	Seed Corn	tembotrione followed by mesotrione

*S-metolachlor + simazine were applied each year before crop and weed emergence



Atrazine resistance due to metabolic detoxification

Typical injury observed in resistant plants at <u>10,000 g/ha</u>12 DAT



Aatrex



Visual injury following atrazine POST (12 DAT) 14.4 kg/ha







Metribuzin (Sencor, Tricor) is still effective on atrazine-resistant waterhemp



Manage weed seedbanks



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HPPD-R waterhemp is not fire-resistant!

(Summer 2011 - greenhouse accident)



Summary and Conclusions

- Dioecious Amaranthus species possess multiple mechanisms for herbicide resistance, including metabolic resistance that mimics corn and cereal crops
- Diverse metabolic enzymes (GSTs and P450s, others?) may be encoded by <u>single</u> or <u>multiple</u> genes
- However, metabolic resistance within the HPPD-inhibitor class in waterhemp can be *herbicide-dependent*
 - cross- or multiple resistance patterns
- Integrated management systems should be utilized



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Dr. Adam Davis and Dr. Aaron Hager, UIUC weed science











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Validation and Implementation of Unmanned Aerial Systems in a Sorghum Breeding Program

N. Ace Pugh, David W. Horne, Geraldo Carvalho Jr., Lonesome Malambo, Anjin Chang, Xiongzhe Han, Jinha Jung, S. Delroy Collins, Sorin C. Popescu, Alex Thomasson, Dale Cope, Murilo M. Maeda, and **William L. Rooney**

Sorghum (Sorghum bicolor, L. Moench) improvement scientists have improved many key characteristics in the crop including biomass yield, disease resistance, and height; however, the rate of improvement is stymied by a phenotyping bottleneck. Unmanned aerial systems (UAS) serve as an attractive potential solution to this problem due to their high temporal and spatial resolution. To that end, several studies have been conducted to determine the ability of rotary-wing and fixed-wing UAS to accurately and efficiently phenotype several traits of interest. First, UAS were evaluated for their ability to estimate plant height in various ideotypes of sorghum. Correlations (r) between rotary-wing UAS and ground-truth measurements ranged from moderate to very high, dependent upon the ideotype of the measured germplasm. In addition, UAS could estimate similar amounts of genetic variance when compared to the ground-truth methodologies, and repeatabilities (R^2) were very high. Another trait of interest was that of anthracnose (Colletotrichum sublineolum) resistance, wherein normalized difference vegetation index (NDVI) estimates were obtained from a fixed-wing UAS to estimate disease incidence and severity. The NDVI served as an excellent predictor of disease severity in later stages of growth, and showed stronger relationships to grain yield than traditional visual scores. Finally, a rotary-wing UAS system could predict biomass yield with a great degree of accuracy, and the relationship between the UAS measurements. An added advantage of UAS is that they enable high-resolution multitemporal growth and progress curves for height, biomass, and disease presence. Using growth curves, sorghum breeders could make novel determinations about their material and discover new phenotypes that were previously difficult to study.

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A field-based high-throughput phenotyping system for tall crops

Maria Salas-Fernandez, Iowa State

High-throughput phenotyping (HTP) technologies have emerged as a consequence of the need to obtain data at large scale, to increase accuracy and repeatability, and to phenotype plants over time for complex traits that could not be characterized by hand. Field-based HTP efforts have focused on the use of unmanned aerial vehicles (UAVs) and the deployment of ground-based platforms carrying sensors or cameras with a top down view on short crops. We have created a novel field-based self-propelled platform equipped with high resolution cameras that was specifically designed for tall crops, to collect images with a side view. This technology has been tested in sorghum and used to obtain plant architecture parameters such as plant height, stem diameter and novel canopy descriptors. The accuracy of image-based algorithmically-derived data was demonstrated when compared with ground-truth measurements. The phenotypic data generated in this project has been used to discover genes/SNPs associated with variation in plant architecture traits and could be further utilized for the genetic improvement of sorghum. The platform and completely automated processing methods developed in our study are new tools for plant breeders and represent significant contributions to the emerging field of predictive phenomics.



Phenotyping for Fungal Resistance in Corn

David A. Hubert Senior Scientist - Plant Management and Phenotyping BASF Plant Science, Research Triangle Park, NC

As ever larger and more complex plant populations are being created and utilized, the need for high throughput screening methods becomes greater. Given the size and cost of these experiments, small mistakes can lead to large consequences. BASF Plant Science is a leader in utilizing high throughput phenotyping for gene discovery and trait characterization. Our focus on imaging and sophisticated sampling has created many learning opportunities to refine techniques and streamline processes. We would like to share our general learnings in working with reverse genetic populations, image analysis, and turning a low throughput assay into a high throughput screen.

Infection by Fusarium species in corn is responsible for yield losses of several hundred thousand bushels of corn annually. Consequently, Gibberella and Fusarium stalk rot are two of the most important diseases in corn. Caused by *Fusarium graminearum* and *Fusarium verticillioides*, respectively, these important diseases are for several reasons possibly the most difficult diseases in corn to study. As such, they also provide an excellent illustrative example of how to move from a very low throughput assay into a high throughput screen.





Outline

- About Phenotyping in BASF Plant Science
- Reverse Genetic Populations and Imaging
- Phenotyping for Fusarium Stalk Rot
- Alternative Bioassay for Maize:Fusarium Interaction
- Primary Bioasssay for Maize:Fusarium Interaction





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Facts about Bioscience Research



Plant Science



Plant Science aims for a better quality of life and an improved environment. We drive innovative solutions for agriculture, nutrition and industrial applications, creating value for BASF and customers.

Plant Science expertise in plant biotechnology is applied to understand crops and to enhance their performance.

Plant Science is focusing on four strong pillars: <u>yield</u> increase, omega-3 fatty acids, herbicide tolerance and <u>fungal resistance</u>. We collaborate with BASF's Operating Divisions or partners such as Monsanto and Cargill.

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About Phenotyping at BASF Plant Science

Focus on Gene Discovery and Better Understanding Traits

Strength in Image Analysis

- Easily automated
- Data can be analyzed many times as new information becomes available
- Highly quantitative for downstream and future analyses

Strong Interactions between Greenhouse and Field Research

Ensures relevance of greenhouse research and good use of resources

High-Throughput Phenotyping and Sophisticated Sampling

Multiple screens running simultaneously each with multiple phenotypes utilizing multiple species studying multiple traits



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Considerations When Working in Reverse Genetic Populations

Issues

- Mutagenesis/ Transformation affects plant growth
- Hidden genetic contributions
- Positional Effects
- Strong Nursery Effects





- Wildtype
- Segregating Null
- Empty Vector
- Efficacious Gene
- Experimental Average
- Phenotypic Extremes
- Mock Treatment

D - BASF

Important Considerations for Image Analysis





Gibberella and Fusarium Stalk and Ear Rot

Maize diseases caused by Fusarium graminearum and Fusarium verticilloides have big impacts

Disease	2012 Yield Loss (millions of bushels)
Fusarium seedling blight	37.5
Gibberella stalk rot	43.9
Fusarium stalk rot	124.6
Fusarium ear rot	91.6
Gibberella ear rot	38.7

Estimated corn yield loss from diseases in the top 22 U.S. corn producing states and Ontario, Canada, in 2012 *Mueller and Wise, Purdue Extension publication, 2014. BP-96-12--W* Yield lossLodging

 Mycotoxin contamination



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Stalk Rot Assay- Practical Aspects for Measuring Infection









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Stalk Rot Assay- Practical Aspects for Measuring Infection















Lead Gene Performance



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BASF We create chemistry

Presented by: Edward Ross, M.S. student in the Department of Crop Sciences – University of Illinois

Nitrogen (N) fertilizers are a major pollutant and input cost of maize (*Zea mays*) production, but their negative effects can be mitigated through the development of cultivars with higher nitrogen use efficiency (NUE). Yield increases due to N fertilizers are primarily attributed to increases in kernel number, a yield component that is determined early in kernel development. Responses to N at this early stage of development are difficult to investigate, due to the complex path of N within the plant and difficulties in precisely manipulating N supply at the developing kernel. To gain more control of N metabolites supplied to the developing kernel, an in vitro kernel culturing system was employed. Hybrid plants from crosses of B73 to Mo17, IHP1, and ILP1 were grown under variable N in the field. Developing kernels were dissected three days after pollination and placed in culture with variable N. B73 X Mo17 kernels were assayed with RNA sequencing and metabolite profiling of free amino acids. Trait and gene expression data were integrated using weighted gene correlation network analysis (WGCNA). A subset of gene modules was found to be highly correlated to free amino acid levels, either in cob or kernel tissue. GO term enrichment analysis of these modules indicates that their members are involved in carbohydrate metabolism, N metabolism, DNA packaging, and protein modification. Additionally, these modules contain genes orthologous to components of an N responsive transcriptional network identified in the Arabidopsis thaliana root. Alleles of these genes containing UniformMu transposon insertions have been obtained from the Maize Genetics Cooperation Stock Center, and are currently being introgressed into various backgrounds. Two of these backgrounds are the IHP1 and ILP1 inbred lines from the Illinois Long Term Selection Experiment (ILTSE) for kernel protein concentration, which represent the extremes of N utilization efficiency in maize.



Presented by: Brian Rhodes, M.S. student in Department of Crop Sciences, University of Illinois

An important component to increasing crop productivity is improving Nitrogen Utilization Efficiency (NUtE). In maize this trait is measured as the ratio of grain yield to accumulated plant N. Enhancing NUtE offers substantial economic and environmental benefits, but little is known about the genetic mechanisms that govern variation for NUtE within maize populations. Our group has conducted high density genetic mapping for NUtE in a hybrid population developed from the intermated B73 X Mo17 recombinant inbred lines (IBMRILs), test crossed to the Illinois High Protein 1 (IHP1) inbred line, which has a superior capacity for N uptake but low NUtE. We identified 9 robust strong effect QTL for NUtE that range in size from 14-9030 Kb and aim to identify causal genetic variants. The largest effect QTL is localized to a 2 Mb region on chromosome 1 containing 23 annotated genes, including the high affinity nitrate transporter NRT1.1 B (GRMZM2G161459). The homolog to the maize NRT1.1 B in rice has been shown to contribute to the variation in nitrogen use between indica and japonica cultivars. A second QTL for grain nitrogen/protein concentration has been localized to a single HVA22-like candidate gene that likely regulates autophagy, a process important for nitrogen remobilization. In addition to analysis of mutant alleles and near-isogenic lines for the QTL interval, we have created transgenic maize inbred lines with grain specific expression of this candidate gene. Preliminary results show an increase in grain protein concentration in both the transgenic inbred background and in F1 ears following hybridization. Genome editing experiments are in progress to further verify the function of candidate genes within our NUtE regions. The results of this project will aid the development of maize hybrids that require lower nitrogen inputs and therefore would reduce costs for farmers and mitigate environmental and health effects associated with high ambient nitrogen levels.

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Tissue Culture and Genome Editing in the Illinois Long Term Selection Experiment

Stephen Jinga, Brian Rhodes, Christine Lucas, Stephen Moose University of Illinois at Urbana-Champaign, Department of Crop Sciences

The Illinois Long Term Selection Experiment is a unique genetic resource for identifying and characterizing genes selected for nitrogen use and protein accumulation in maize. To facilitate study of gene functions, we aim to establish a CRISPR Cas9 mediated genome-editing system in these novel genetic backgrounds. A media regime has been developed for successfully regenerating fertile plants of both Illinois High Protein (IHP) and Illinois Low Protein (ILP). Currently, putative transgenic lines expressing the Cas9 protein have been recovered using NPTII as a selectable marker. These Cas9 positive lines will be used to make targeted mutations with this germplasm. We have also initiated experiments to edit the Prolamin Box Binding Factor (PBF), which regulates zein gene expression and shows changes in both allele frequencies and mRNA expression that are consistent with PBF being a target of selection for grain protein concentration. In addition to generating knockout mutations, we are also investigating the functional significance of variation in the length of an asparagine (Asn) repeat motif found at the C-terminus of PBF. This Asn repeat shares features with triplet repeat expansions studied in Arabidopsis and trinucleotide repeat disorders in humans such as Huntington's disease. It is hypothesized that variation in the Asn rich region of PBF could act as a sensor to control α -zein accumulation in response to incoming supply of amino acids, or possibly interacting with other transcription factors such as opaque-2. To target this Asn-repeat motif, single-guide RNAs were designed to create variation in Asn repeat length in conjunction with expressed Cas9.



Response to Selection in the ILTSE and a population of epigenetic NILs

Nicholas Heller and Stephen Moose Graduate Research Fellow, Department of Crop Sciences University of Illinois at Urbana-Champaign

Advances in sequence technology have allowed incredible discoveries about the genomes of many organisms and elucidated relationships between gene and phenotype. These advances have allowed a deeper look into how variation in phenotype is created, selection for this variation is realized, and how phenotypes are inherited. However, many studies found that the heritability of many phenotypes is not fully explained by genomic DNA sequence, especially for quantitative, complex traits.

Here, I present some insights into the contributions to phenotypic response to selection in plants using a genetic system (utilizing the Illinois Long Term Selection Experiment, ILTSE), transgenic system (utilizing the red fluorescent protein driven by the *Floury2* promoter in the maize kernel), and an epigenetic system (utilizing variation created by the *mop1* mutation). Briefly, the ILTSE materials provide a unique resource because they have undergone continuous selection for the same trait for over 100 years and the last 50 years of seed is preserved. Reverse selection experiments are still underway to determine the plasticity of the populations' genomes after 50, 90, and even 100 years of forward selection. Finally, we utilize an inbred system to look more closely at the possibility that some of the response to selection is due to heritable, non-genetic factors.

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Analysis of habituation at the maize r1 locus

Authors: Robert Lindsay¹, William Eggleston, Jr.²

1. Integrated Life Sciences, Virginia Commonwealth University

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Abstract:

A mutation of the maize rI locus in W22 with variable kernel color was used as the basis for directed selection to create sublines with kernel color ranging from colorless to nearly fully colored. These sublines were produced by 5-6 generations of selecting and planting the lightest and/or darkest kernels on selfed ears from the prior generation. Visual inspection indicates that there are multiple gradations in kernel color among the selected sublines ranging from nearly colorless to nearly fully colored, which is supported by quantification with light reflectometry. However, light reflectometry does not support the full range of color gradations discernable by visual means. Recombination studies indicate that the initial kernel color change was caused by a change in the 3' end (or beyond) of the r1 gene controlling kernel color. Initial sequence analysis of the 3' end of the r1 in the progenitor, lightest subline, and darkest subline does not show sequence changes that could account for the change in seed color differences between the progenitor and two sublines in this region. The lack of sequence differences suggests that the change in kernel color between the sublines are due to epigenetic changes, rather than DNA sequence changes, and may result from a process known as "habituation." resulting from a lack of canalization at the r1 locus. This idea of habituation, supported by methylation changes in the selected sublines holds promise for reducing the time required to isolate and develop crop phenotypes.


Determining the effect of the *sbe1* allele from Z. mays parviglumis on maize endosperm starch composition in an *ae1* background

P. Awale and D. Auger

Department of Biology and Microbiology, South Dakota State University, Brookings, SD.

Starch is the main constituent of maize endosperm. Structurally, starch is divided between two main forms: unbranched (or less branched) amylose and highly branched amylopectin. Generally, amylose constitutes about 25% of maize endosperm starch. The amylose content in the endosperm is increased up to 50% when *ae1*, which encodes starch branching enzyme IIb (SBEIIb), is homozygous recessive. However, one variety of maize that is homozygous *ae1*, GEMS-0067, has up to 75% amylose in its endosperm starch. We have shown that this high amylose content is due to an allele of *sbe1*, which encodes for starch branching enzyme I (SBEI). The GEMS-0067 allele of *sbe1* translates into a protein with six amino acid polymorphisms relative to what is found in all Midwestern dents that have been surveyed. We have also found that the amino acid sequence for SBEI from GEMS-0067 is identical to what is predicted for *Z. mays parviglumis*. We are interested in whether the *sbe1* allele of *Z. mays parviglumis* has the same effect on maize starch composition as GEMS-0067. To test this, we will analyze the progeny of *Z. mays parviglumis*-maize hybrids. Instead of using a recessive *ae1* allele, we are employing *Ae1-5180*, which acts in a dominant fashion to eliminate SBEIIb. We will present our methods of analysis as well as data on developing markers to distinguish the *sbe1* and *ae1* alleles.

Keywords: Amylose, ae1, sbe1, Z. mays parviglumis, GEMS-0067



Mapping loci that modify the efficacy of *Teosinte crossing barrier 1*

Merritt B. Burch and Donald Auger

Teosinte crossing barrier 1 (Tcb1) is a genetic cross-incompatibility factor that is responsible for blocking non-self-type pollen in silks. Originally found in teosintes, *Tcb1-s* (strong allele) has been introduced into modern maize varieties conferring resistance to tcb1 pollen. Previous studies using a similar cross incompatibility system, Gametophye factor 1 (Ga1-s) suggest that the cell wall modification enzyme ZmPme3, a pectin methylesterase, along with multiple modifying QTL loci contribute to the effectiveness of silks at resisting foreign pollen types. In Tcb1, little is known about the genetic modifiers and, more importantly, what the underlying biological mechanism is for this cross incompatibility. Cross-incompatibility systems like Tcb1 and Ga1 can be beneficial to breeders and farmers when only certain pollen types are desired on specialty maize crops. It was observed that nearly all the F1's of various inbreds, including B73, crossed by W22 Tcb1-s demonstrate strong incompatibility with tcb1 pollen. One exception was Mo17, whose F1s had weaker resistance. In this study we used recombinant inbred lines (RILS) from the intermated B73-Mo17 (IBM) population crossed with homozygous W22 Tcb1-s plants to test the efficacy of the various F1s at blocking tcb1 pollen. The F1s were tested by first challenging the Tcb1-s silks with R1 C1 tcb1 pollen and the next day pollinated the same silks with r1 c1 Tcb1-s pollen. The resulting ears were scored for the percentage of colored kernels. Six quantitative trait loci (QTL) were detected on chromosomes 1, 3, 5, and 7 that explained 28.9% of the phenotypic variability. Most modifying QTL loci showed simple additivity effects and epistatic interactions between loci. Further exploration into these genomic regions and the underlying candidate genes is underway, these results could shed light on the genetic and physiological mechanisms controlling *Tcb1*.



Interaction plot for csu207 and umc1752





Error 70 18.74110 0.267730 Total 76 32.80192

Drop one QTL at a time ANOVA table:

	df	Туре	III SS	LOD	%var	F	value	Pvalue(Chi2)	<pre>Pvalue(F)</pre>	
1@28.0	1		2.0279	1.7179	6.182		7.575	0.005	0.00753	**
1@324.0	1		0.4868	0.4288	1.484		1.818	0.160	0.18186	
3@21.0	1		1.2024	1.0397	3.666		4.491	0.029	0.03762	*
3@107.0	1		1.9384	1.6456	5.909		7.240	0.006	0.00891	**
5@426.0	1		1.2296	1.0626	3.749		4.593	0.027	0.03558	*
7@4.0	1		2.5974	2.1702	7.918		9.702	0.002	0.00267	**
Signif.	cod	les:	0 ****	0.001	د**،	0.0)1'*'	0.05 '.' 0.1	'' 1	



Crop Sciences college of agricultural, consumer & environmental sciences Deployment of a High-Throughput Plant Height Mapping System on Genomes to Fields Germplasm

Jessica Bubert, M.S. student Department of Crop Sciences, University of Illinois at Urbana-Champaign

A remote sensing system for crop height measurement was developed using a 360-degree 2D laser scanner and onboard computer mounted on an unmanned aerial vehicle (UAV). Algorithms for data processing and visualization were developed to process the dense data generated by the Light Detection and Ranging system (LiDAR) on the UAV. This system was deployed on a two acre field of various maize inbreds and hybrids in nitrogen use experiments, including the Genomes to Fields germplasm. Plant heights for each plot were generated from the resulting spatial point clouds. Across the field, the plant heights determined via remote sensing show an average R² value of 0.87 when compared to manual plant height measurements. In the Genomes to Fields experiment plant heights were taken at tassel height and at the point of flag leaf attachment. The comparison of these to the LiDAR data will be used to determine where on the plant the LiDAR system is recognizing a point for the spatial point cloud. Future research can determine if late season fly-overs to develop a spatial point cloud of a field post-leaf senescence could provide a high-throughput method of measuring ear height in addition to plant height. This UAV-based system can cover a two-acre field in just eight and a half minutes, providing a truly high-throughput and non-destructive measure of plant height that can be used throughout the growing season and across locations.



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